Synthesis of 2-Amino-2,5,6-trideoxy-5-phenylphosphinyl-L-galactopyranose. The P-in-the-Ring Analogue of L-Fucosamine

Tadashi Hanaya, Hiroshi Yamamoto, Heizan Kawamoto, Margaret-Ann Armour,†
Alan M. Hogg,† and Hiroshi Yamamoto*

Department of Chemistry, Faculty of Science, Okayama University, Tsushima, Okayama 700

† Department of Chemistry, The University of Alberta, Edmonton, Alberta, T6G 2G2, Canada (Received April 17, 1992)

3-O-Benzyl-1,2-O-isopropylidene-6-O-tosyl- α -D-allofuranose was led to methyl (5R and 5S)-3-O-benzyl-5,6-dideoxy-5-[(methoxy)phenylphosphinyl]-D-ribo-hexofuranoside (9) in a 4 step sequence (55% overall yield). Methyl (5R and 5S)-2-acetamido-2,5,6-trideoxy-5-[(methoxy)phenylphosphinyl]-D-arabino-hexofuranoside (12) was prepared from 9 in 4 steps involving a C-2 inversion (62% overall yield). Compound 12 was then converted, in 2 steps, into the title compound, which was characterized as the peracetyl derivatives.

We have prepared various sugar analogues having a phosphorus atom in the hemiacetal ring¹⁾ because of a significant interest in their chemical and biochemical properties. Thus, a large number of P-in-the-ring sugar analogues were synthesized, such as those of D-ribofuranose 1,20 D-glucopyranose 2,30 and 6-deoxy-L-galactopyranose (L-fucose) 3.4) However, no such analogues of an amino sugar type have been reported yet, whereas some 2-amino-2,6-dideoxyhexoses have long been known to occur in nature; for instance, L-fucosamine (4) has been identified as a component of the antigenic lipopolysaccharide of the Pseudomonas species.⁵⁾ wish to describe herein a convenient synthesis of 2amino-2,5,6-trideoxy-5-phenylphosphinyl-L-galactopyranose (a P-in-the-ring analogue of 4) as the first example of an amino sugar type by employing phenylphosphinyl as a model functional group.

Treatment of 3-O-benzyl-1,2-O-isopropylidene-6-O-

tosyl- α -D-allofuranose (5)⁶⁾ with pyridinium chlorochromate (PCC) gave the 5-ulose derivative 6 (Scheme 1). The reaction of 6 with methyl phenylphosphinate in 1,2-dimethoxyethane in the presence of DBU yielded 5,6-

Scheme 1.

anhydro-5-[(methoxy)phenylphosphinyl] compound 7. Hydrogenation of 7 in the presence of Raney Ni (W-4) in ethanol afforded the 5,6-dideoxy derivative 8.

Compound 8 was treated with methanol in the presence of acidic ion-exchange resin to give methyl glycoside 9. It was then led to the 2-O-trifluoromethanesulfonate to introduce an azido group with C-2 inversion, affording methyl D-arabino-hexofuranoside 10. Azide 10 was easily led to the 2-acetamido derivative 11 upon treatment with thiolacetic acid or by hydrogenation in the presence of 10% Pd on carbon, followed by acetylation. Debenzylation of 11 by catalytic hydrogenation over 20% Pd(OH)₂-C provided furanoside 12.

Then, compound 12 was reduced with sodium dihydrobis(2-methoxyethoxy)aluminate (SDMA), followed by acid hydrolysis, to afford a mixture of 2,5,6-trideoxy-2-amino-5-phenylphosphinyl-L-galactopyranose (13) and D-altropyranose (14). These were converted into their tetraacetyl derivatives, which were purified by column chromatography over silica gel with 19:1 (v/v) ethyl acetate-ethanol, thus affording 2-acetamido-1,3,4-tri-O-acetyl-2,5,6-trideoxy-5-[(R)-phenylphosphinyl]- α -L-galactopyranose (15a, 7.2% overall yield from 12), its β -anomer 15b (5.2%), 2-acetamido-1,3,4-tri-O-acetyl-2,5,6-trideoxy-5-[(S)-phenylphosphinyl]- α -D-altropyranose (16a, 1.0%), its β -anomer 16b (5.5%), and 5-[(R)-P]- α -isomer 16c (1.6%).

The precise structures of 15a, b and 16a—c were established by analysis of their 500-MHz 1 H NMR spectra; for all the assignments of the signals, see Table 1. The $^{1}C_{4}$ conformation of 15a, b and 16c are derived from the large

values of $J_{4,P}$ (26—29 Hz) and $J_{2,3}$ (ca. 11 Hz) and small values of $J_{2,P}$ (0—4 Hz).⁴⁾ In contrast, the opposite magnitudes ($J_{4,P}$ =ca. 4, $J_{2,3}$ =3—5, $J_{2,P}$ =24—27 Hz) of **16a**, **b** suggest the 4C_1 conformation. The small $J_{5,P}$ values for **15a**, **b** and **16a**, **b** indicate the axial orientation of H-5 and the ring P=O group, 3,4) whereas the equatorial H-5 orientation of **16c** is assigned by the large $J_{5,P}$ value. Since a similar argument can be applied to $J_{1,P}$ values, the anomeric orientation at C-1 is readily perceived by these values as well as those of $J_{1,2}$. It is noteworthy that a considerable downfield shift (δ =1.6—2.0) is observed for the signals of axial HN-2 group (for **16a**, **b**) in the vicinity of axial P=O group compared with those of equatorial HN-2 group (for **15a**, **b** and **16c**).

Although improvement of the yields of the final ring enlargement reactions and isolation of other minor isomers of 15 and 16 remain to be done, the present work demonstrates a convenient way for preparation of the P-in-the-ring analogue of 2-amino-2,6-dideoxyhexoses and this scheme is expected to be readily applicable for preparation of various other amino sugar analogues.

Experimental

Melting points were determined with a Yanagimoto MP-S3 instrument and are uncorrected. All reactions were monitored by TLC (Merck silica gel 60F, 0.25 mm) with an appropriate solvent system [(A) 1:1 AcOEt-hexane, (B) 19:1 AcOEt-EtOH, or (C) 19:1 CHCl₃-MeOH]; components were detected by exposing the plates to UV light and/or by spraying them with 20% sulfuric acid-ethanol, with subsequent heating. Column chromatography was performed by Wako C-200 silica gel. The IR spectra were taken with a Hitachi 260-10S

Table 1. ¹H and ³¹P NMR Parameters for 15a, b and 16a—c in CDCl₃

| Compd | Chemical shifts/δ | | | | | | | | | | | | |
|-------|-----------------------|--------------------|-----------|-----------|---------------|------|------|--------------------|--------------|--------------------|-----------|--------------------|--------------------|
| | H-1 | H-2 | H-3 | H-4 | H-5 | H-6 | NH-2 | | Ac-1,2,3,4a) | | | $Ph(o,m,p)^{b)}$ | |
| 15a | 5.70 | 5.36 | 5.30 | 5.71 | 2.66 | 1.17 | 5.50 | 2. | 27, 2.06, 1 | .92, 1.91 | 7.73, | 7.51, 7.63 | 29.2 |
| 15b | 5.58 | 5.11 | 5.31 | 5.64 | 2.31 | 1.14 | 5.53 | 2. | 26, 2.05, 1 | .97, 1.91 | 7.77, | 7.52, 7.59 | 29.8 |
| 16a | 5.16 | 4.84 | 5.45 | 5.63 | 2.98 | 1.12 | 7.17 | 2. | 16, 2.08, 2 | 2.03, 1.97 | 7.79, | 7.56, 7.65 | 38.9 |
| 16b | 5.78 | 4.85 | 5.57 | 5.54 | 2.55 | 1.08 | 7.49 | 2. | 23, 2.10, 2 | 2.03, 1.92 | 7.75, | 7.57, 7.65 | 37.0 |
| 16c | 5.96 | 5.22 | 5.41 | 5.42 | 2.71 | 1.05 | 5.47 | 2. | 26, 2.08, 2 | 2.00, 1.93 | 7.73, | 7.52, 7.59 | 32.3 |
| Compd | Coupling constants/Hz | | | | | | | | | | | | |
| | $J_{1,2}$ | $J_{1,\mathrm{P}}$ | $J_{2,3}$ | $J_{2,P}$ | $J_{2, m NH}$ | ı J | 3,4 | $J_{3,\mathrm{P}}$ | $J_{4,5}$ | $J_{4,\mathrm{P}}$ | $J_{5,6}$ | $J_{5,\mathrm{P}}$ | $J_{6,\mathrm{P}}$ |
| 15a | 2.5 | 10.5 | 11.4 | 0 | 8.7 | 2 | .6 | 0 | 2.6 | 27.6 | 7.3 | 6.5 | 14.8 |
| 15b | 11.7 | 3.3 | 10.8 | 4.4 | 8.9 | 2 | .5 | 0 | 2.5 | 28.9 | 7.1 | 3.0 | 14.4 |
| 16a | 2.9 | 9.1 | 3.0 | 24.3 | 8.5 | 3 | .0 | 1.0 | 11.9 | 4.2 | 7.1 | 4.5 | 15.5 |
| 16b | 4.8 | 3.2 | 4.7 | 27.0 | 8.5 | 2 | .6 | 1.5 | 12.0 | 3.6 | 7.0 | 3.1 | 15.1 |
| 16c | 11.4 | 3.0 | 10.6 | 3.5 | 9.6 | 2 | .5 | 0 | 4.0 | 26.1 | 8.0 | 19.4 | 16.4 |

a) The assignment of acetyl signals may be interchanged. b) J values for P-Ph: $J_{P,o}=12.5$, $J_{P,m}=3.5$, $J_{P,p}=1.5$, $J_{o,m}=J_{m,p}=7.5$ and $J_{o,p}=1.5$ Hz.

infrared spectrophotometer. The ¹H and ³¹P NMR spectra were measured in CDCl₃ with Varian VXR-500 and VXR-200 instruments (500 and 81 MHz, respectively, the SC-NMR Lab., Okayama Univ. at 21°C) or with a Hitachi R-1900 instrument (90 MHz, FT). Chemical shifts are reported as δ values relative to tetramethylsilane (internal standard for ¹H) and 85% phosphoric acid (external standard for ³¹P). The mass spectra were taken on an A.E.I. MS 50 ultrahigh resolution instrument and were given in terms of m/z (rel intensity) compared with the base peak.

3-O-Benzyl-1,2-O-isopropylidene-6-O-tosyl-α-D-ribo-hexofuranos-5-ulose (6). To a suspension of $5^{(6)}$ (1.10 g, 2.38 mmol) and finely powdered molecular sieves 3A (2.4 g) in dry CH₂Cl₂ (20 ml) was added PCC (1.54 g, 7.14 mmol) at 0°C. The mixture was stirred at 20°C for 6 h and then 2-propanol (6 ml) was added at 0°C. The mixture was stirred at 20°C for 1 h and diluted with ether. The precipitates were filtered off through celite. The filtrate was evaporated in vacuo and the residue was purified by short-path column chromatography with 1:1 AcOEt-hexane as an eluant, giving 6 (0.99 g, 90%) as a colorless syrup: $R_1 = 0.59$ (A); IR (neat) ν 1740 (C=O), 1350 (ν_{as} SO₂), 1170 cm⁻¹ (ν_s SO₂); ¹H NMR (500 MHz) δ =1.35, 1.57 (3H each, 2s, CMe₂), 2.43 (3H, s, MeC₆-S), 3.84 (1H, dd, $J_{3,4}$ =9.1, $J_{2,3}$ =4.2 Hz, H-3), 4.52 (1H, t, $J_{1,2}$ =3.5 Hz, H-2), 4.53 (1H, d, H-4), 4.64, 4.71 (1H each, 2d, J=12.0 Hz, CH₂O-3), 4.84, 4.86 $(1 \text{H each}, 2 \text{d}, J_{6,6} = 17.7 \text{ Hz}, \text{H-}6,6'), 5.77 (1 \text{H}, \text{d}, \text{H-}1), 7.33, 7.81$ $(2H \text{ each}, 2d, J=8.1 \text{ Hz}, C_6H_4-S), 7.32-7.37 (5H, m, Ph).$

(5R and 5S)-5,6-Anhydro-3-O-benzyl-1,2-O-isopropylidene-5-[(R and S)-(methoxy)phenylphosphinyl]- α -D-ribo-hexofuranoses (7). To a solution of 6 (2.89 g, 6.24 mmol) and methyl phenylphosphinate (1.24 ml, 9.34 mmol) in DME (20 ml) was dropwise added a solution of DBU (0.98 ml, 6.57 mmol) in DME (7.0 ml) at -40 °C. The mixture was stirred at the same temp for 1 h and then concentrated in vacuo. The residue was dissolved in CH₂Cl₂ and saturated aq NaHCO₃ was added. After having been stirred at 20°C overnight, the organic layer was separated and the aq layer was extracted twice with CH₂Cl₂. The combined organic layers were dried (Na₂SO₄) and evaporated in vacuo. The residue was chromatographed in a column of silica gel, giving 7 (2.43 g, 87%) as a colorless syrup: R_1 =0.15 (A); ³¹P NMR δ =32.1, 33.1, 35.2, and 36.1 (in a ratio of 18:35:31:16, respectively); ¹H NMR (500 MHz) for the two major components of 7 δ =1.32, 1.35,* 1.44, 1.58* (6H, 4s, CMe₂), 2.95,* 2.97 (1H, d,* dd, $J_{6,P}$ =4.9,* 4.1, $J_{6,6}$ =5.7 Hz, H-6'), 2.95,* 3.26 (1H, d,* t, $J_{6,P}$ =4.9,* 4.8 Hz, H-6), 3.63, 3.74* $(1H, 2dd, J_{3,4}=8.8, 8.5, *J_{2,3}=4.4 Hz, H-3), 3.78, *3.80 (3H, 2d,$ J_{POMe} =10.8 Hz, POMe), 4.30, 4.59* (1H, 2d, $J_{4,P}$ =2.5, 4.2* Hz, H-4), 4.42, 4.48,* 4.63, 4.71* (2H, 4d, J=11.3 Hz, CH₂O-3), 4.51, 4.54* (1H, 2td, $J_{1,2}=3.7$, ${}^{5}J_{2,P}=1.5$ Hz, H-2), 5.70,* 5.71 (1H, 2d, H-1), 7.30—7.57 [8H, m, Ph-C, Ph(m,p)-P], 7.83,* 7.91 [2H, m, $J_{o,P}$ =12.2, $J_{o,m}$ =7.1, $J_{o,P}$ =1.4 Hz, Ph(o)-P], for the two minor components of 7 δ =1.29, 1.36,* 1.46, 1.56* (6H, 4s, CMe₂), 2.33, 3.01* (1H, 2dd, $J_{6'P}$ =6.2, 2.6,* $J_{6,6'}$ =5.2, 5.6* Hz, H-6'), 2.87, 3.26* (1H, 2t, $J_{6,P}$ =5.2, 4.8* Hz, H-6), 3.56, 3.78* $(3H, 2d, J_{POMe} = 10.8 \text{ Hz}, POMe), 3.75 = 3.80 (1H, m, H-3), 3.91,$ $4.57*(1H, dd, m, *J_{4,P}=15.1, J_{3,4}=8.1 Hz, H-4), 4.48, 4.57*(1H, dd, m, *J_{4,P}=15.1, J_{4,P}=15.1, J_{4,P}=15.1,$ 2m, H-2), 4.56, 4.57, * 4.71, 4.74* (2H, 4d, J=12.0 Hz, CH₂O-3), 5.21, 5.86* (1H, 2d, $J_{1,2}$ =3.4 Hz, H-1), 7.30—7.57 [8H, m, Ph-C, Ph(m,p)-P, 7.71, 7.79* [2H, m, Ph(o)-P].

(5R and 5S)-3-O-Benzyl-5,6-dideoxy-1,2-O-isopropylidene-5-[(R and S)-(methoxy)phenylphosphinyl]- α -D-ribo-hexofuranoses (8). Compound 7 (832 mg, 1.86 mmol) dissolved in ethanol (8 ml) was hydrogenated in the presence of Raney-Ni (W-4) (1.65 g) at 20°C under an atmospheric pressure of H₂. After 8 h, the catalyst was filtered off through celite, and the filtrate was evaporated in vacuo. The residue was chromatographed on silica gel to give 8 (555 mg, 69%) as a colorless syrup: $R_1 = 0.11 (A)$; ¹H NMR (90 MHz) $\delta = 0.97$, 1.01 (3H, 2dd, $J_{6,P}$ =16.5, $J_{5,6}$ =7.3 Hz, H₃-6), 1.25, 1.29, 1.32, 1.37, 1.48, 1.50 (6H, 6s, CMe₂), 2.20—2.50 (1H, m, H-5), 3.59, 3.64, 3.66, 3.67 $(3H, 4d, J_{POMe}=11.8 \text{ Hz}, POMe), 3.90-4.35 (2H, m, H-3.4),$ 4.40—4.80 (3H, m, H-2, CH₂O-3), 5.53, 5.68, 5.74 (1H, 3d, $J_{1,2}$ =3.5 Hz, H-1), 7.33 (5H, m, Ph-C), 7.35—7.80 (5H, m, Ph-P); ${}^{31}P$ δ =46.4, 35.7, 45.1 (33:31:35); MS m/z 432 (M+; 0.4), 341(29), 326(7), 309(6), 283(15), 213(24), 184(33), 155(51), 91(100). Found: m/z 432.1709. Calcd for $C_{23}H_{29}O_6P$: M, 432.1703.

Methyl (5R and 5S)-3-O-Benzyl-5,6-dideoxy-5-[(R and S)-(methoxy)phenylphosphinyl]-α(and β)-D-ribo-hexofuranosides (9). A mixture of 8 (455 mg, 1.05 mmol) and Amberlite IR-120B (H⁺) ion exchange resin (2 ml) in abs methanol (8 ml) was refluxed for 2 h. After cooling, the resin was filtered off and the filtrate was evaporated in vacuo. The residue was purified by column chromatography with a gradient eluant of CHCl₃ \rightarrow 5% MeOH-CHCl₃, giving 9 (407 mg, 95%) as a colorless syrup: R_i =0.40—0.36 (C); ¹H NMR (90 MHz) δ=1.07, 1.10, 1.25 (3H, 3dd, $J_{6,P}$ =17.4, $J_{5,6}$ =7.3 Hz, H₃-6), 2.05—2.45 (1H, m, H-5), 2.80—3.20 (1H, m, HO-2), 3.02, 3.17, 3.20, 3.33 (3H, 4s, MeO-1), 3.60, 3.62, 3.64, 3.66 (3H, 4d, J_{POMe} =11.7 Hz, POMe), 3.75—4.40 (3H, m, H-2,3,4), 4.50—4.71 (3H, m, H-1, CH₂O-3), 7.30—7.85 (5H, m, Ph-P), 7.33 (5H, m, Ph-C).

Methyl (5R and 5S)-2-Azido-3-O-benzyl-2,5,6-trideoxy-5-[(R and S)-(methoxy)phenylphosphinyl]- α (and β)-D-arabino-hexofuranosides (10). To a solution of 9 (377 mg, 0.928 mmol), 4dimethylaminopyridine (6.0 mg, 0.05 mmol) and dry pyridine (0.30 ml, 3.7 mmol) in CH₂Cl₂ (5 ml) was added trifluoromethanesulfonic anhydride (0.30 ml, 1.8 mmol) at -40°C over a period of 30 min. The stirring was continued at this temp for 1 h and then water (0.5 ml) was added. The organic layer was separated and aq layer was extracted with CH₂Cl₂. combined organic layers were dried (Na₂SO₄) and evaporated in vacuo. The residue was dissolved in DMF (2.5 ml) and sodium azide (300 mg, 4.6 mmol) was added. The mixture was stirred at 40°C for 36 h. The precipitate was filtered off and the filtrate was concentrated in vacuo. The residue was dissolved in CH2Cl2 and washed with water, dried (Na2SO4) and evaporated in vacuo. The residue was chromatographed on silica gel using 5% methanol-chloroform as the eluant, giving **10** (295 mg, 74%) as a colorless syrup: R_f =0.40 (C); IR (neat) ν 2100 cm⁻¹ (N₃); ¹H NMR (90 MHz) δ =0.98, 1.04, 1.23, 1.48 $(3H, 4dd, J_{6,P}=16.3, J_{5,6}=7.5 Hz, H_3-6), 2.05-2.45 (1H, m, H-6)$ 5), 3.03, 3.20, 3.32, 3.33, 3.35 (3H, 5s, MeO-1), 3.60, 3.62, 3.63, 3.65, 3.68 (3H, 5d, J_{POMe} =11.0 Hz, POMe), 3.70—3.95 (1H, m, H-3), 3.95—4.40 (2H, m, H-2,4), 4.50—4.95 (3H, m, H-1, CH₂O-3), 7.30—7.85 (5H, m, Ph-P), 7.33 (5H, m, Ph-C); MS m/z 400 (M-CH₃O; 0.6), 316(3), 251(25), 155(42), 91(100). Found: m/z 400.1426. Calcd for $C_{20}H_{23}N_3O_4P$: M-CH₃O, 400.1426.

Methyl (5R and 5S)-2-Acetamido-3-O-benzyl-2,5,6-trideoxy-5-[(R and S)-(methoxy)phenylphosphinyl]- α (and β)-Darabino-hexofuranosides (11). A. A mixture of azide 10 (456 mg, 1.06 mmol) and thioacetic acid (0.47 ml, 6.6 mmol) was stirred

^{*} For another diastereomer with regard to the phosphorus atom; the assignment of some of the δ values may have to be interchanged.

at room temp for 30 h under argon. The excess thioacetic acid was removed under reduced pressure. The resulting syrup was chromatographed in a column of silica gel, giving 11 (411 mg, 87%) as a colorless syrup: R_f =0.33-0.28 (C). Recrystallization of this mixture from AcOEt-hexane gave one pure diastereomer as colorless prisms (62 mg): Mp 190-191°C; ¹H NMR (500 MHz) δ =1.21 (3H, dd, $J_{6,P}$ =16.4, $J_{5,6}$ =7.3 Hz, H₃-6), 1.99 (3H, s, AcN-2), 2.28 (1H, dquint, $J_{5,P}=14.7$, $J_{4,5}=$ 7.0 Hz, H-5), 3.15 (3H, s, MeO-1), 3.64 (3H, d, J_{POMe} =10.9 Hz, POMe), 3.94 (2H, m, H-3,4), 4.51, 4.71 (1H each, 2d, J=11.1 Hz, CH₂O-3), 4.58 (1H, ddd, $J_{2,NH}$ =8.8, $J_{2,3}$ =6.1, $J_{1,2}$ =4.9 Hz, H-2), 4.71 (1H, d, H-1), 6.06 (1H, d, NH-2), 7.31 (5H, m, Ph-C), 7.43 [2H, m, $J_{o,m}=J_{m,p}=7.5$, $J_{m,p}=3.3$ Hz, Ph(m)-P], 7.51 [1H, m, $J_{o,p}=J_{p,P}=1.4 \text{ Hz}, \text{ Ph}(p)-P$], 7.75 [2H, m, $J_{o,P}=11.4 \text{ Hz}, \text{ Ph}(o)-$ P]; ${}^{31}P$ NMR $\delta=45.5$; MS m/z 416 (M-CH₃O; 0.3), 356 (M-PhCH₂; 0.2), 341(3), 324(5), 296(7), 254(10), 184(28), 155(34), 91(100). Found: m/z 416.1621. Calcd for $C_{22}H_{27}$ -NO₅P: M-CH₃O, 416.1627.

B. Compound **10** (33 mg, 0.076 mmol) dissolved in ethanol (0.3 ml) was hydrogenated in the presence of 10% Pd-C (40 mg) at 20°C under an atmospheric pressure of H₂. After 4 h, the catalyst was filtered off through celite, and the filtrate was evaporated in vacuo to give 2-amino-2-deoxy derivative as a colorless syrup. This was dissolved in dry pyridine (0.06 ml) and acetic anhydride (0.03 ml) was added at 0°C. After stirring at 20°C overnight, cold water was added. The mixture was evaporated in vacuo and the residue was chromatographed on silica gel to give **11** (28 mg, 82%).

Methyl (5R and 5S)-2-Acetamido-2,5,6-trideoxy-5-[(R and S)-(methoxy)phenylphosphinyl]- α (and β)-D-arabino-hexofuranosides (12). Compound 11 (110 mg, 0.246 mmol) dissolved in ethanol (2 ml) was hydrogenated in the presence of 20% Pd(OH)₂–C (80 mg) at room temp under an atmospheric pressure of H₂. After 6 h, the catalyst was filtered off through celite, and the filtrate was evaporated in vacuo. The residue was purified by short-path column chromatography to give 12 as a colorless syrup (87 mg, 99%): R_i =0.15 (C); ¹H NMR (90 MHz) δ =0.92, 1.11, 1.22, 1.28 (3H, 4dd, $J_{6,P}$ =16.5, $J_{5,6}$ =7.5 Hz, H₃-6), 1.97, 1.98, 2.00 (3H, 3s, AcN-2), 2.05—2.45 (1H, m, H-5), 3.20, 3.25, 3.30 (3H, 3s, MeO-1), 3.25—3.50 (1H, m, HO-3), 3.59, 3.64, 3.69 (3H, 3d, J_{POMe} =11.0 Hz, POMe), 3.70—4.25 (2H, m, H-3,4), 4.25—4.80 (2H, m, H-1,2), 6.10—6.35 (1H, m, NH-2), 7.35—7.80 (5H, m, Ph-P).

2-Acetamido-1,3,4-tri-O-acetyl-2,5,6-trideoxy-5-[(R and S)phenylphosphinyl]- α (and β)-L-galactopyranoses (15) and D-Altropyranoses (16). To a solution of 12 (192 mg, 0.537 mmol) in dry benzene (3 ml) was added, with stirring, a solution of SDMA (3.4 M in toluene, 0.75 ml, 4.8 equiv) in dry benzene (2 ml), in small portions at 5°C under argon. The mixture was stirred at this temp for 2 h, and then water (0.5 ml) was added. The mixture was centrifuged and the precipitate was extracted with several portions of benzene. The organic layers were combined and evaporated in vacuo. The residue was immediately treated with 1:1 2-propanol-0.5 M hydrochloric acid (6.0 ml) (1 M=1 mol dm⁻³) at 90°C for 3 h under argon. After cooling, the reactants were neutralized with Amberlite IRA-45. The resin was filtered off and washed with aq ethanol. The filtrate was evaporated in vacuo to give a mixture of 2-amino-2,5,6-trideoxy-5-[(R and S)-phenylphosphinyl]- α (and β)-Lgalactopyranoses (13) and D-altropyranoses (14) as a colorless syrup.

This was dissolved in dry pyridine (4.0 ml) and acetic anhydride (2.0 ml) at 0° C. The mixture was stirred at 20° C overnight, diluted with a small amount of cold water, and concentrated in vacuo. The residue was dissolved in CH_2Cl_2 and washed with water. The organic layer was dried (Na_2SO_4) and evaporated in vacuo, giving a mixture of tetraacetates 15 and 16, which were separated by column chromatography with a gradient eluant of $AcOEt \rightarrow 19:1$ AcOEt - EtOH into three fractions A-C.

Fraction A [R_i =0.36 (B)] gave a colorless syrup (15.3 mg) which consisted of 5-[(S)-phenylphosphinyl]- α -D-altropyranose **16a** (1.0% from **12**) and its 5-[(S)-P]- β -altro isomer **16b** (5.5%), the relative amounts being determined by the integral ratio of ³¹P signals: ¹H and ³¹P NMR, see Table 1: MS m/z 440 (M+1; 3), 397(7), 380(58), 338(17), 320(36), 277(48), 259(100), 236(21), 195(27), 125(77). Found: m/z 440.1473. Calcd for $C_{20}H_{27}NO_8P$: M+1, 440.1473.

Fraction B $[R_i=0.21 (B)]$ gave 5-[(R)-P]- α -L-galacto isomer **15a** as a colorless syrup: 17.0 mg (7.2% from **12**): 1 H and 31 P NMR, see Table 1; MS m/z (M+; 7), 396(23), 380(100), 338(32), 320(66), 277(65), 259(43), 236(36), 195(36), 125(79). Found: m/z 439.1405. Calcd for C_{20} H₂₆NO₈P: M, 439.1396.

Fraction C $[R_i=0.10 \ (B)]$ gave a colorless syrup (16.0 mg) which consisted of 5-[(R)-P]- β -L-galacto isomer **15b** (5.2% from **12**) and 5-[(R)-P]- β -D-altro isomer **16c** (1.6%): ¹H and ³¹P NMR, see Table 1; MS m/z 439 (M+; 12), 397(38), 380(50), 338(41), 320(48), 277(78), 251(89), 236(45), 203(77), 125(100). Found: m/z 439.1398. Calcd for $C_{20}H_{26}NO_8P$: M, 439.1396.

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References

- 1) H. Yamamoto and T. Hanaya, "Studies in Natural Products Chemistry," ed by T. I. Atta-ur-Rahman, Elsevier, Amsterdam (1990), Vol. 6, pp. 351—384; H. Yamamoto and S. Inokawa, Adv. Carbohydr. Chem. Biochem., 42, 135 (1984).
- 2) T. Hanaya and H. Yamamoto, *Bull. Chem. Soc. Jpn.*, **62**, 2320 (1989); H. Yamamoto, Y. Nakamura, S. Inokawa, M. Yamashita, M.-A. Armour, and T. T. Nakashima, *J. Org. Chem.*, **49**, 1364 (1984).
- 3) T. Hanaya, A. Akamatsu, H. Kawamoto, M.-A. Armour, A. M. Hogg, and H. Yamamoto, *Bull. Chem. Soc. Jpn.*, **64**, 2398 (1991); H. Yamamoto, T. Hanaya, H. Kawamoto, S. Inokawa, M. Yamashita, M.-A. Armour, and T. T. Nakashima, *J. Org. Chem.*, **50**, 3516 (1985); H. Yamamoto, K. Yamamoto, S. Inokawa, M. Yamashita, M.-A. Armour, and T. T. Nakashima, *ibid.*, **48**, 435 (1983).
- 4) T. Hanaya, H. Yamamoto, T. Ohmae, H. Kawamoto, M.-A. Armour, A. M. Hogg, and H. Yamamoto, *Bull. Chem. Soc. Jpn.*, **64**, 869 (1991).
- 5) G. T. Barry and E. Roark, *Nature*, **202**, 493 (1964); J. A. Cifonelli, P. A. Rebers, M. B. Perry, and J. K. N. Jones, *Biochem.*, **5**, 3066 (1966).
- 6) G. W. J. Fleet, N. G. Ramsden, R. A. Dwek, T. W. Rademacher, L. E. Fellows, R. J. Nash, D. St. C. Green, and B. Winchester, *J. Chem. Soc., Chem. Commun.*, **1988**, 483.